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Exploring anticoagulant rodenticide exposure and effects in eagle owl (*Bubo bubo*) nestlings from a Mediterranean semiarid region

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ABSTRACT

Anticoagulant rodenticides (ARs) are widely used for pest control, resulting in their pervasive presence in the environment and posing significant toxicological risks to a range of predatory and scavenging species. Our study mainly aimed to evaluate AR exposure and effects in nestlings of eagle owl (Bubo bubo) from the Region of Murcia (southeastern Spain). We analysed ARs in blood samples (n = 106) using high-performance liquid chromatography-triple quadrupole (HPLC-TQ), assessed the influence of potential anthropogenic (presence of livestock farms, landfills and human population density) and environmental (land uses and proximity to watercourses) variables, and measured prothrombin time (PT) and plasma biochemical parameters as biomarkers of effects. Our results showed the presence of AR residues in 91.5% of the nestlings, with 70.8% exhibiting multiple ARs (up to six compounds in a single individual). Second-generation ARs (SGARs) were the most prevalent compounds. The analysis of biochemical parameters indicated that the sampled individuals were in good physiological condition. Although PT was positively correlated with total AR concentration (SARs), the relationship was not significant (Rho = 0.04; p = 0.49). Regarding environmental factors, higher ΣARs were associated with the most urbanised study site and the presence of landfills, likely due to the increased availability of rodent prey. The prevalence of two SGARs (brodifacoum and difenacoum) was linked to closer proximity to riverbeds, suggesting a contamination pathway associated with inland aquatic ecosystems, where these AR compounds may concentrate due to water scarcity. This study underscores the widespread exposure of eagle owls to ARs and highlights the importance of effective monitoring and management of these pollutants to protect conservation-concern wildlife in Mediterranean semiarid regions.

1. Introduction

The natural habitat of birds of prey is increasingly being taken over by human activities, leading to a variety of conservation issues. Urban expansion and agricultural intensification contribute to the degradation and fragmentation of their habitats and to the loss of nesting sites (Mainwaring, 2015; McClure et al., 2018). In addition, the widespread use of chemicals, particularly rodenticides and other pesticides, introduces toxic substances into the environment that can contaminate their food sources, leading to bioaccumulation and adverse health effects (Badry et al., 2021; González-Rubio et al., 2021; Shore and Taggart, 2019). Moreover, raptors often inhabit wildland-urban interface areas, where natural habitats intersect with human development (Radeloff et al., 2005), increasing their vulnerability to environmental hazards resulting from human activities.

A significant problem affecting raptors is their frequent and

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widespread exposure to anticoagulant rodenticides (ARs) (García-Fernández et al., 2024; Gomez et al., 2022; Nakayama et al., 2019; Ruiz-Suárez et al., 2014). Birds of prey are particularly at risk because their feeding habits often include target rodents, and their biological characteristics make them more susceptible to the toxic effects of these compounds (Khidkhan et al., 2024; Nakayama et al., 2020; Rattner et al., 2012). It is therefore crucial to understand the extent of AR exposure, both in terms of spatial distribution and severity, in order to implement mitigation measures where necessary, particularly for species that are declining and for which AR contamination could be a significant risk factor (Spadetto et al., 2024b; Vicedo et al., 2024).

In an effort to mitigate the impact of these compounds on non-target wildlife, the European Union applied new regulations in 2018, classifying AR products exceeding concentrations of 30 ppm as reprotoxic (European Union, 2016). Consequently, in Spain and other countries of the European Union, the products available to the general public now contain lower concentrations, often involving second-generation ARs (SGARs), which are more potent and persistent in the environment than first-generation ARs (FGARs) (Erickson and Urban, 2004). Interestingly, it has recently been shown that AR baits with lower concentrations (25 ppm) were more palatable and therefore more consumed than those with higher concentrations (50 ppm). As a result, target rodents carried higher hepatic AR levels at the time of their death (Frankova et al., 2024). While the effects of these compounds at low doses on non-target fauna are still not fully understood, recent studies indicate that AR exposure and concentrations remain high in non-target predatory birds (Carrillo-Hidalgo et al., 2024; Moriceau et al., 2022; Spadetto et al., 2024b).

The Eurasian eagle owl (*Bubo bubo*, hereafter referred to as eagle owl) is a top predator with a diet primarily consisting of potentially pest species in the study area, such as European rabbits (*Oryctolagus cuniculus*) and rats. This large nocturnal raptor has a wide distribution across the Palearctic region, and it is monitored over the long term in numerous countries, which facilitates the replication of toxicological assessments on a large scale. Additionally, the eagle owl is considered a species of conservation interest, as it is included in the List of Species in Special Protection Regime (LESPRE) (Gobierno de España, 2011) and Annex I of the Birds Directive (European Union, 2009). With a high ecological plasticity in habitat selection, it breeds near highly anthropized areas such as intensive crops or landfills, potentially exposing it to ARs. Moreover, the ease of studying its diet and the abundant literature on its trophic ecology allow for a more thorough understanding of the spatiotemporal patterns in AR exposure.

The Region of Murcia, located in the southeast of the Iberian Peninsula, features a mix of agricultural lands, rural villages, and cities, creating a unique interface where wildlife habitats are increasingly intersected by human development. This region hosts high densities of eagle owls in areas where its primary prey, the European rabbit, is abundant. In areas with lower rabbit availability, the eagle owl diet diversifies, with species like hedgehogs, rats, and birds becoming more significant (León-Ortega et al., 2016; Pérez-García et al., 2012). Given its high territorial density in the study area, the eagle owl has already been used as a biomonitoring species for the presence of heavy metals and other environmental contaminants (Espín et al., 2014; Gómez-Ramírez et al., 2012a,b).

Previous studies have already highlighted the presence of ARs in various avian species across the southeastern Iberian Peninsula (Gómez-Ramírez et al., 2021; Spadetto et al., 2024b; Vicedo et al., 2024), particularly in populations breeding near human settlements, likely due to domestic use of these substances. The eagle owl occupies diverse environments, including territories located in natural areas as well as near potential exposure sources (e.g., livestock farms, landfills or human settlements). Additionally, this species often breeds and hunts in rocky gorges and along riverbeds, where its main prey is more abundant. However, the role of watercourses as pathways for AR exposure in top predators has been scarcely evaluated in semi-arid regions. Here,

non-perennial watercourses could serve as collection points for run-off containing pesticides and toxic chemicals (Arenas-Sánchez et al., 2016), while also acting as vital water sources for wildlife during dry periods (Steward et al., 2012). As a consequence, the potential for biomagnification of ARs through the food chain increases. Additionally, recent studies suggest that raptors in semi-arid regions may depend on free water sources, particularly during droughts (Boal et al., 2023; O'Brien et al., 2006), which could further amplify their exposure to contaminants. Therefore, we set out to assess the influence of these riparian ecosystems on AR exposure, along with the presence of irrigation reservoirs near the breeding territories.

The aims of this study were 1) to evaluate AR exposure in eagle owl nestlings in a semi-arid Mediterranean landscape, 2) to determine if there was a relationship between potential risk factors (land uses, livestock farms, human population density, landfills and riverbed presence) with AR prevalence and concentration in blood, and 3) to assess the usefulness of a coagulation parameter as a biomarker of AR effect and study a plasma biochemical profile to evaluate the general health status of the chicks. Our hypotheses were that AR exposure could be widespread in this species, and that the rate of AR exposure is higher in eagle owls exposed to the considered risk variables, compared to those inhabiting more natural or protected areas. Additionally, we hypothesised that the coagulation capacity decreases proportionally to the levels of ARs detected in blood samples, as previously demonstrated in other bird of prey species (Spadetto et al., 2024a, 2024b). By providing critical insights into the extent of AR contamination and its impact on the eagle owl, this study seeks to provide information for the development of conservation strategies and regulatory measures to mitigate the risks posed by these chemicals.

2. Material and methods

2.1. Study area

The Region of Murcia is characterised by a typically warm and dry Mediterranean climate. Summer temperatures can usually reach 40 °C, while winters are mild with average temperatures around 10–15 °C. Precipitation is scarce, with an annual average of less than 300 mm (Spanish National Agency of Meteorology - AEMET, 2024). The region is known for its intensive cultivation of citrus fruits, vegetables, and orchards, highlighting its importance as an agricultural area. In fact, in 2022 agricultural land occupied 32% of the entire region, with 47% of irrigation agriculture (CARM Región de Murcia, 2022).

Within the region, eagle owl territories of our study were distributed across four geographic zones (Fig. 1), each with distinct geographic and environmental features.

- Central-eastern badlands, referring to a badland area geologically characterized by extensive erosion and lack of vegetation, where non-perennial rivers supply water to the region. This study site surrounds the city of Murcia and it is characterized by a significant human presence, with a dense network of small settlements, rural houses, and agricultural structures scattered throughout the landscape. It hosts a high density of eagle owls, also due to the high availability of European rabbits.
- 2) Prelittoral mountain ranges, predominantly occupied by a regional park (Carrascoy y El Valle), along with a Site of Community Importance (SCI) and a Special Protection Area for Birds (SPA), which partially overlap. These areas are characterized by an arid mountainous landscape with a variety of rocky and hilly formations and are mainly covered by Mediterranean vegetation (Aleppo pine forests and scrubland). However, in some flat areas or wider valleys, limited agricultural activities such as fruit tree cultivation can be found. Human presence in these areas is limited, with few scattered settlements. The presence of eagle owls here is also considerable.



Fig. 1. Eagle owl territories (triangles) sampled in the Region of Murcia (southeastern Spain) in 2021–2022 for the purpose of assessing exposure to anticoagulant rodenticides (ARs). The main municipalities of the region are marked with a star.

- 3) Western badlands, within the municipality of Lorca. Agriculture in these areas may be limited due to soil conditions and water availability, but it is still present with rainfed crops including olive groves, almond trees, and cereal crops. The human population is generally sparse, with small and dispersed settlements throughout the territory. This area shows a high density of eagle owl breeding territories.
- 4) Littoral mountain ranges, an arid and mountainous terrain located south of an extensive coastal plain (Campo de Cartagena) and overlooking the coast of the Mediterranean Sea, with its climate and ecosystem influenced by the proximity to the sea. Human presence is mainly concentrated in urban centres such as Cartagena and La Unión, while the rest of the surrounding territory is characterized by small villages and scattered farms. The density of eagle owls here is much lower than in other areas, as is the presence of the European rabbit. Hence, they mainly feed on other prey items, predominantly rats (Sánchez-Virosta et al., 2020).

2.2. Sample collection

Sampling was conducted as part of a nocturnal raptor monitoring and marking program in the Region of Murcia. Territories were carefully monitored from December to June to estimate the egg laying, hatching, and fledging dates of the chicks and to choose the appropriate time to access the nests. Nests were accessed by specialized personnel once the chicks were approximately 30–45 days old, before reaching the fledging age. A total of 106 blood samples from eagle owl nestlings were collected during the breeding season of 2021 (n = 39) and 2022 (n =67). Owlets belonged to a total of 34 territories, of which 10 were sampled in both study years, while the remaining 24 were sampled only once due to territorial occupation, either in 2021 or 2022, for a total of 44 sampling events.

The sample collection (approved by the Ethical Committee for

Animal Experimentation of the University of Murcia; code 657/2020) was carried out following a protocol for monitoring contaminants in raptors (Espín et al., 2021). Blood samples were extracted from the brachial vein using a sterile syringe with a 23G needle and transferred into a heparinized tube (2 mL). In 2022, an aliquot of 450 μ L was transferred to a tube with 50 μ L of 0.109 M sodium citrate buffer. The samples were kept refrigerated until arrival at the laboratory, which occurred within a few hours. Then, an aliquot of 1 mL of heparinized blood (n = 105) was centrifuged at 2500 g for 10 min to obtain plasma, while the samples preserved in sodium citrate data samples. All the samples were stored at -80 °C until the time of analysis.

2.3. Anticoagulant rodenticide analysis

Blood samples were analysed to detect the presence of 10 ARs (bromadiolone, brodifacoum, chlorophacinone, coumatetralyl, difenacoum, diphacinone, flocoumafen, coumafuryl, and warfarin), using the method outlined by Spadetto et al. (2024b). In summary, 1000 µL of acetonitrile and 25 μ L of coumachlor (internal standard) were added to 250 µL of blood, in a tube with a ceramic homogeniser, followed by vortexing the tube for 1 min. Next, extraction salts, comprising 0.25 g of NaCl and 1 g of Na_2SO_4 per sample, were introduced. The tube was manually shaken for 1 min and then centrifuged at 2500 g for 5 min. The resulting supernatant was collected and transferred to a tube containing purification products (12.5 mg of PSA, 37.5 mg of C18, and 225 mg of Na₂SO₄). After vortexing for an additional minute, the tube was centrifuged at 2500 g for 5 min. The supernatant was then drawn using a syringe and transferred into a chromatography vial after filtering through a 0.45 µm nylon syringe filter. Detailed information on chemicals and standards can be found in Box 1 SI. The extracts underwent analysis for the aforementioned ARs using an HPLC-TQ system

(consisting of vacuum degasser, autosampler and a binary pump; Agilent Series 1260, Agilent Technologies, Santa Clara, CA, USA) equipped with a reversed phase C18 analytical column (150 \times 2.1 mm and 2.6- μm particle size; Phenomenex Kinetex R 2.6 µm EVO Polar C18 100 A) and an Ultivo G6303 triple quadrupole mass spectrometer from Agilent, with an electrospray ionisation interface, following the procedure described by Spadetto et al. (2024b). The values of the SRM ratios from sample extracts for all of the two transitions selected were between 6% and 21% of average of calibration standards from same sequence. This is in accordance with the Analytical quality control and method validation procedures for pesticide residues analysis in food and feed (SANTE/11312/2021 v2; European Commission, 2021), based on ion-ratio statistics for the transitions monitored. Under the chromatographic conditions described above, the calibration graphs were constructed by plotting peak area vs. concentrations in the range 0.1–50 ng mL^{-1} . The limits of quantification (LOQ), calculated following the guidance in the document SANTE/11312/2021 v2 (European Commission, 2021), ranged between 0.01 and 2.5 ng mL⁻¹. The analytical technique exhibited recovery values between 76% and 105%, with a relative standard deviation < 14%. This complies with the recovery and precision accepted by the SANTE/11312/2021 v2 (European Commission, 2021).

2.4. Coagulation assays and biochemical analysis

The coagulation tests were conducted using a coagulometer, as detailed in Spadetto et al. (2024b). Fibrinogen was measured to assess the quality of the sample, as a decrease in its levels (less than 50–60 mg dL⁻¹; Rattner et al., 2010, 2020) may indicate improper sample collection or handling, as well as potential liver or kidney diseases or other pathologies. Briefly, a kit from Spinreact S.A.U (Spain) based on the Clauss method (Clauss, 1957) was utilized. Citrated plasma samples were diluted 1:10 with imidazole buffer. After that, 200 μ L of the dilution were combined with 20 μ L of kaolin in a tube containing a mixer. After incubating the tube for 3 min at 37 °C, 100 μ L of bovine thrombin were added, and the clot formation time was measured.

Prothrombin time (PT) was chosen as an indicator of AR effect on blood coagulation, as it rapidly alters following the ingestion of ARs and has been previously used for this purpose (Hindmarch et al., 2019; Rattner et al., 2010, 2011, 2014a). To ensure reliable results in avian species, avian thromboplastin should be used as a reagent. In fact, commercially available kits typically contain mammalian thromboplastin, leading to inaccurate results (reviewed by Webster, 2009). The thromboplastin was obtained using the Quick method modified by Griminger et al. (1970), as described in Spadetto et al. (2024b). For the PT test, 50 mg of chicken thromboplastin were reconstituted in 2.5 mL of CaCl₂ in a Falcon tube. The mixture was agitated with a mixer for 15 min and centrifuged at 1800 rpm for 20 min. The supernatant was diluted 1:1 with CaCl2 and 200 µL of the prepared reagent were added to a tube with a mixer and incubated for 3 min at 37 °C. Subsequently, 100 µL of citrated plasma were added to initiate the reaction, and the coagulometer measured the clotting time.

All analyses of biochemical parameters in plasma samples were performed using an Olympus A400 biochemical analyser with commercially available reagents (Beckman Co), with the exception of ovotransferrin, which was measured using an ELISA kit (Chicken Ovotransferrin ELISA Kit, ab157694). All assays had intra- and inter-assay imprecision <15% and were linear after serial dilutions.

2.5. Statistical analysis

Some environmental variables were selected and assessed as potential drivers of AR exposure. These factors were calculated within 1-km radius buffers constructed around each nest using QGIS Geographic Information System open source (version 3.32.3). The buffer radius was chosen as an approximation of the eagle owl's home range in the study area, as reported in the literature (León-Ortega, 2016; Pérez-García et al., 2012). Land use data were extracted from the CORINE Land Cover 2018; EEA, 2018), supplemented with information from the SIOSE Land Use Map (IGN, 2016) to provide a more detailed land use categorization. To allow for a more reliable analyses, land uses were classified into the five main classes: artificial areas, agricultural land, natural vegetation, wetlands, and water bodies. Additional subclasses were calculated within the "agricultural land" category: total irrigated and non-irrigated crops and the percentage of land occupied by irrigated orchards. In order to evaluate the role of the hydrological network in AR contamination, we calculated the minimum distance of each nest to the nearest watercourse (using a hydrological network layer; Gobierno de España, 2024), and the number of irrigation reservoirs within the buffer. Moreover, the human population density of the census section where the nest is located was used, extracting data from the latest population census conducted in 2021 (National Institute of Statistics, 2021). Finally, livestock farm data for 2021 in the Region of Murcia were provided by the Spanish Ministry of Agriculture, Fisheries, and Food, allowing for the calculation of animal density (i.e., of pigs, sheep and goats, poultry, equines, and total livestock) as well as the total number of farms within the buffer. Poultry and equines were excluded from the analysis, as they were present in very low numbers near the sampled territories. The variables used are described in detail in Table 1 SI.

Descriptive statistics were computed for each compound, as well as for the sum of SGARs, FGARs, the 10 analysed ARs (ΣARs), PT and biochemical parameters. To examine the association between the selected environmental factors (refer to Table 2 SI) with the concentration and prevalence of ARs, we utilized the information-theoretic method pioneered by Burnham and Anderson (2002). Linear Mixed Models (LMM) were constructed using the "lme" function from the "nlme" package (Pinheiro et al., 2023), treating environmental variables as fixed effects and territory as a random factor. Model comparisons were based on the bias-corrected version of Akaike's information criterion (AICc). Models were ranked using AICc differences (Δ AICc) and Akaike weights (w). \triangle AICc was calculated as the difference between the AICc of each model and the AICc of the best model. Models with \triangle AICc <2 can be alternative models to the selected model. Akaike weights may be interpreted as the probability that a given model is the actual best model of the set (Burnham and Anderson, 2002). To compare the prevalence and concentration of Σ ARs across years, we employed the "glmer" function from the "lme4" package (Bates et al., 2015), with a logit link and binomial error distribution, regarding the breeding territory as a random factor.

Spearman's correlation test was employed to evaluate the relationships between Σ ARs and PT and between Σ ARs and the plasma biochemical parameters. All statistical analyses were conducted using R software version 4.3.1, and significance levels were set at p < 0.05.

3. Results

3.1. AR levels and prevalence

The data resulting from the two years of study were grouped and analysed together since no significant interannual differences were found in either AR prevalence or Σ ARs. At least one AR was detected in 91.5% of the samples (n = 106). The prevalence of FGARs was markedly lower (11.3%) compared to that of SGARs (91.5%). All the analysed compounds were detected, except coumafuryl. The compound with the highest prevalence was flocoumafen (79.2%), followed by difenacoum (49.1%), brodifacoum (41.5%), and bromadiolone (28.3%) (Table 1). Considering the breeding territory as a unit of analysis, compounds were detected in 90.9% of the sampling events (40/44).

Multiple ARs were detected in 70.8% of the nestlings. Indeed, in individuals testing positive, one to six compounds were detected (Fig. 2). The total AR concentration (Σ ARs) in positive individuals ranged from 0.03 to 57.81 ng mL⁻¹ (with a median of 0.77 ng mL⁻¹).

Table 1

Prevalence (%) and levels (ng mL⁻¹) of anticoagulant rodenticides (ARs) detected in eagle owl nestlings (n = 106) sampled in the Region of Murcia (southeastern Spain). Descriptive statistics are provided for individuals with detected levels of ARs (expressed as n+ in the table).

	$\mathbf{n}+$	%	Mean	Median	SD	Min.	Max.
FGARs							
Chlorophacinone	2	1.9	0.24	0.24	0.15	0.13	0.34
Coumafuryl	0	0	-	-	-	-	-
Coumatetralyl	4	3.8	0.14	0.08	0.14	0.04	0.34
Diphacinone	2	1.9	0.60	0.60	0.45	0.28	0.92
Warfarin	5	4.7	0.02	0.02	0.01	0.01	0.04
SGARs							
Brodifacoum	44	41.5	0.72	0.27	1.63	0.06	9.87
Bromadiolone	30	28.3	0.39	0.24	0.44	0.03	2.09
Difenacoum	52	49.1	1.34	0.22	7.02	0.05	50.83
Difethialone	4	3.8	0.74	0.65	0.49	0.26	1.40
Flocoumafen	84	79.2	1.26	0.16	6.32	0.02	57.43
ΣFGARs	12	11.3	0.19	0.08	0.26	0.01	0.92
ΣSGARs	97	91.5	2.34	0.74	7.81	0.03	57.81
ΣARs	97	91.5	2.37	0.77	7.81	0.03	57.81



Fig. 2. Ring plot showing the percentage of eagle owl nestlings in the Region of Murcia (southeastern Spain) for which varying numbers of anticoagulant rodenticide (AR) compounds were detected. The number at the top of each segment indicates the specific number of AR compounds found, including cases where no compounds (0) were detected.

Coinciding with the prevalence, the compounds yielding the highest values were flocoumafen (57.43 ng mL⁻¹), difenacoum (50.83 ng mL⁻¹), and brodifacoum (9.87 ng mL⁻¹).

3.2. Analysis of factors that may influence AR exposure

Based on statistical analyses, it appears that none of the selected variables were related to AR prevalence (understood as the overall prevalence of all ARs analysed collectively) (Table 3 SI). Applying the same model to the prevalence of SGARs with the highest detection rates (flocoumafen, bromadiolone, difenacoum, and brodifacoum), we found that brodifacoum and difenacoum were associated with the distance from the nearest river (Table 4 SI and 5 SI; Fig. 4). Sheep-goat and swine density were slightly higher than the null model in the case of flocoumafen and bromadiolone, respectively (Table 6 SI and 7 SI). However, these models differ only slightly from the null model. Similarly, for



Fig. 3. Effect of different study sites on total concentration of anticoagulant rodenticides (Σ ARs) in eagle owls (*Bubo bubo*) nestlings from the Region of Murcia (southeastern Spain). The plot shows concentration estimates (ng mL⁻¹) and 95% confidence intervals.

brodifacoum, two variables (livestock density and study site) were above the null model (Table 4 SI), but the difference was minimal and thus not truly explanatory of the detected brodifacoum prevalence.

Regarding AR levels, these were related to the study site (the highest ranked variable) and the proximity to a landfill (Table 2). Indeed, the highest concentrations of ARs were detected in the Central-eastern badlands study site (Fig. 3). The remaining variables considered did not prove to be explanatory of the observed Σ AR concentrations.

3.3. Coagulation tests and biochemical analysis

The fibrinogen level was found to be above 100 mg dL⁻¹ in all analysed samples (range 108.9–589.2 mg dL⁻¹), indicating that the samples were collected and handled correctly and that the individuals examined had sufficient fibrinogen levels to ensure normal blood coagulation. PT in eagle owl nestlings averaged 13.03 ± 1.83 s (range 8.8–17.5 s). A pair of values was considered an outlier and excluded from the analysis (57.81 ng mL⁻¹; 12.5 s). Although positive, no significant relationship was detected between Σ ARs and PT (*Rho* = 0.04; *p* = 0.49), as shown in Fig. 1 SI.

Regarding the analysis of biochemical parameters in plasma samples, descriptive statistics were calculated, excluding a single ALT value, considered an outlier due to its significantly higher level (373 UI L⁻¹). Results are presented in Table 3. Σ ARs was not found to be correlated with any of the plasma biochemical parameters analysed, except for glucose (*Rho* = 0.225; *p* = 0.029).

4. Discussion

As mentioned above, the eagle owl is considered an ideal biomonitoring species in the study area (Gómez-Ramírez et al., 2021). However, as far as we know, this is only the second biomonitoring study of AR exposure using blood samples from eagle owl nestlings. In the first study, Gómez-Ramírez et al. (2012b) analysed 50 blood samples from free-living eagle owls (both adults and nestlings) from southeastern Spain, but no individuals tested positive. However, it is possible that the technique used was not sufficiently sensitive to detect low levels of ARs.

The presence of ARs in the eagle owl has been predominantly studied



Fig. 4. The plots show the effect of the distance to the nearest watercourse on difenacoum (a) and brodifacoum (b) prevalence in eagle owl (*Bubo bubo*) nestlings in the Region of Murcia (southeastern Spain). Shadow areas represent 95% confidence intervals.

Table 2

Ranking of the factors used to explain the total concentration of ARs (Σ ARs) detected in nestling eagle owls (*Bubo bubo*) in the Region of Murcia (south-eastern Spain).

	k	AICc	ΔAICc	w
Study site	6	725.745	0.000	0.796
Landfill	4	729.201	3.456	0.141
Null model	3	731.968	6.223	0.035
Farm number	4	734.245	8.500	0.011
Irrigation reservoirs	4	735.647	9.902	0.006
Artificial areas	4	736.646	10.901	0.003
Orchards	4	738.466	12.721	0.001
Non irrigated crops	4	738.516	12.771	0.001
Irrigated crops	4	738.676	12.931	0.001
Natural vegetation	4	739.029	13.284	0.001
Agricultural land	4	739.157	13.412	0.001
Human density	4	740.947	15.202	0.000
Swine density	4	744.633	18.888	0.000
Sheep-goat density	4	745.658	19.913	0.000
Watercourse distance	4	748.845	23.100	0.000
Livestock density	4	749.750	24.005	0.000

k = number of parameters estimated; AICc = corrected Akaike's Information Criterion; $\Delta AICc$ = difference between AICc of each model and the minimum AICc; w = Akaike's weight.

using liver samples from carcasses found in various European countries, although the number of samples is generally low (i.e. < 20 individuals). Nonetheless, the detection rate is notably high, ranging from 50 to 100% (Christensen et al., 2012; Fourel et al., 2024; Langford et al., 2013; López-Perea et al., 2015, 2019; Moriceau et al., 2022; Sánchez-Barbudo et al., 2012). In fact, AR residues were detected in 83% of liver samples from adult individuals (n = 18) found in an area adjacent to ours (Gómez-Ramírez et al., 2021). Additionally, Fourel et al. (2024) recently found that the median sum of SGARs in eagle owls (180 ng g^{-1} ww, n =14) was the highest among the species from France and Réunion Island examined in the study. Similarly, Christensen et al. (2012) found a substantial median concentration (241 ng g⁻¹ ww, n = 10) in eagle owl liver samples from Denmark, a value closely approaching the highest concentration recorded in the red kite (*Milvus milvus*; 260 ng g⁻¹, n = 3). In López-Perea et al. (2015), 64.3% of the individuals (n = 14) from Catalonia (Spain) had hepatic concentrations of SGARs higher than 200 ng g^{-1} ww. Such concentrations exceed the assumed toxicity level for ARs (100–200 ng g⁻¹, Newton et al., 1999; Thomas et al., 2011) and are therefore considered potentially lethal. These data are significant because they do not only highlight that the eagle owl is a species easily exposed to ARs, but also may show higher capacity than other species to accumulate these compounds, which increases the risk of adverse effects.

Overall, AR concentrations detected here were low and fell within the range found in other studies conducted on blood samples for nestlings of other birds of prey (Badry et al., 2022; Martínez-Padilla et al., 2017; Oliva-Vidal et al., 2022; Powolny et al., 2020; Spadetto et al., 2024b). However, two of our owlets presented unusually elevated Σ AR concentrations, exceeding 50 ng mL⁻¹ (50.9 and 57.8 ng mL⁻¹). These AR levels have been the highest detected so far in blood samples from free-living raptor nestlings, and presumably indicate the recent consumption of a prey with high concentrations of SGARs (flocoumafen and difenacoum respectively). These data are quite alarming, as they suggest that episodes of exposure to high AR doses might be occurring more frequently than expected, with possible acute or subacute effects in the short term.

The prevalent compounds were SGARs, as expected, due to their greater potency and effectiveness compared to FGARs. They are also preferred for their ability to overcome the widespread resistance that rodent populations have developed against FGARs. Flocoumafen was also detected at very high prevalence in other nocturnal and diurnal raptors in the Region of Murcia sampled during the same study years (Spadetto et al., 2024a, 2024b). These results are remarkable given that, in Spain, only one product containing flocoumafen is registered for non-professional use, and two are intended for use by specialized personnel (Carrera et al., 2024). Difenacoum and brodifacoum were also detected with very high prevalences, which likely indicates their widespread use in the area, possibly due to the large number of registered products for both professional and non-professional use (69 for difenacoum and 129 for brodifacoum) (Carrera et al., 2024). The low prevalence of difethialone (3.8%) suggests that it may be the only second-generation compound scarcely used in the study area. FGAR diphacinone is not authorized for use in Europe (EC Regulation 528/2012) (European Union, 2012), yet it was detected in two eagle owl nestlings. Similar findings were reported for other raptor (Vicedo et al., 2024) and mammal (Carrera et al., 2024) species sampled in the same area (SE Spain), suggesting illegal use of this compound.

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Table 3

Descriptive statistics (median, mean, SD, minimum and maximum) of biochemical parameters in the plasma of eagle owl nestlings (n = 105) from the Region of Murcia (southeastern Spain).

	Median	Mean	SD	Minimum	Maximum
Creatinine (mg dL ⁻¹)	0.36	0.39	0.19	0.05	1.83
Total proteins (g dL^{-1})	3.14	2.90	0.64	1.05	4.30
Albumin (g dL^{-1})	1.21	1.22	0.10	0.95	1.52
Calcium (mg dL ⁻¹)	9.77	10.04	2.46	8.35	34.27
Amylase (UI L ⁻¹)	529.40	516.54	121.93	175.00	810.70
Creatine kinase (UI L^{-1})	1516.10	1616.39	772.50	268.40	3817.80
Cholesterol (mg dL ⁻¹)	199.11	202.57	34.93	127.16	291.47
Alkaline phosphatase (UI L ⁻¹)	600.20	623.53	140.00	330.40	1250.60
Phosphorus (mg dL^{-1})	7.14	7.12	0.79	5.24	9.03
γ -glutamyltransferase (UI L ⁻¹)	4.40	5.19	4.85	0.70	43.70
Glucose (mg dL^{-1})	409.90	413.26	27.62	357.20	523.60
Aspartate aminotransferase (UI L ⁻¹)	225.60	244.04	92.42	113.30	748.70
Alanine aminotransferase (UI L^{-1})	37.60	43.94	36.31	11.00	373.00
Urea (mg dL $^{-1}$)	13.70	14.33	6.27	4.00	45.20
Total bilirubin (mg dL^{-1})	0.10	0.11	0.04	0.03	0.39
Uric acid (mg dL^{-1})	12.99	13.88	4.57	5.47	25.42
Magnesium (mg dL $^{-1}$)	2.34	2.42	0.45	1.75	4.96
Lactate dehydrogenase (UI L^{-1})	882.60	969.17	453.90	292.00	2304.50
Ovotransferrin (μ g mL ⁻¹)	12137.00	12607.46	6058.91	1155.00	34400.00
Butyrylcholinesterase (mol $mL^{-1} min^{-1}$)	1.10	1.17	0.33	0.10	2.60
Acetylcholinesterase (mol $mL^{-1} min^{-1}$)	1.76	1.81	0.38	0.30	3.14

4.1. Variables affecting AR exposure

Significantly higher ΣAR concentrations have been detected in the study site of the Central-eastern badlands, where the level of urbanisation near eagle owl territories is the highest among the study sites in the region. Eagle owls in our study area often reside in the wildland-urban interface (zones where urban or residential development is directly adjacent to or mixed with natural vegetation), where it has been recently shown that the risk of AR exposure can be significant (Hofstadter et al., 2021; Silveira et al., 2024). In another research conducted on common kestrels in the same study area, higher Σ SGAR levels have been observed in the more urbanised study site, surrounding the city of Murcia (Spadetto et al., 2024a), which suggests that in heavily anthropized areas, the residential use of ARs is widespread among the local population. These findings are also consistent with other studies in which the degree of urbanisation has been found to be a strong predictor of AR exposure (Geduhn et al., 2015; Lohr, 2018; López-Perea et al., 2019; Musto et al., 2024). It is worth noting that in the other study sites, particularly in the western badlands and littoral mountain ranges, the number of nests is notably lower, which could be a limitation of our study.

Furthermore, we identified a correlation between Σ AR levels and the proximity of landfills and waste treatment facilities to eagle owl territories. Waste dumps serve as environments with an abundant food supply, attracting numerous wildlife species, particularly rodents. The consequent proliferation of these animals often necessitates the increased use of ARs for population control (Berny et al., 2014; Coeurdassier et al., 2018). As a result, eagle owls that consume these AR contaminated rodents are likely exposed to elevated doses of these compounds. Avian scavengers, such as kites and vultures, often exploit these facilities and were also reported to bioaccumulate ARs in their bodies (Badry et al., 2022; López-Perea et al., 2019; Oliva-Vidal et al., 2022). These findings underscore the critical need for rigorous monitoring of AR application near landfills and the consideration of alternative management strategies to mitigate the impact on non-target raptor and scavenger species.

On the other hand, AR detection in blood essentially indicates recent exposure, as the compounds have a short-lived presence in the blood-stream (e.g., 16.5 ± 10.0 h for warfarin in Eastern barn owl *Tyto javanica*, Khidkhan et al., 2024). We do not know the exact timing of the exposure nor the initial dose. Although it is unclear whether some degree of bioaccumulation in the blood might occur, higher concentrations

in a particular study site could indicate repeated exposures or at least massive application of these substances in the considered area. It is also likely that these products are not correctly applied (e.g., due to lack of bait boxes, incorrect amounts, use in inappropriate areas, and lack of post-application monitoring) and that dead rodents are not promptly removed. Additionally, the prevalence of ARs was high (\geq 75%) in all four study sites and could not be related to any of the environmental variables considered, indicating that, regardless of any factor taken into account, and even in nests located in natural and protected areas, the risk for the eagle owl of being exposed to ARs remains considerable.

Interestingly, the higher likelihood of finding certain SGAR compounds (brodifacoum and difenacoum) in territories near watercourses can be attributed to a combination of ecological, behavioural, and anthropogenic factors. ARs can be transported from application areas to watercourses through surface runoff, especially during rains or irrigations (Regnery et al., 2018). Once in watercourses, they can be further transported downstream and accumulate in nearby areas. Zones adjacent to water bodies, particularly around ponds or riverbeds in semiarid environments, tend to harbour greater biodiversity and rodent density (Shenbrot et al., 2010; Williams, 2005; Zamora-Marín et al., 2021), exposing them to AR contamination. Furthermore, these areas may experience intensive use of ARs to control rodent populations, especially in agricultural zones or near human settlements. Predatory birds such as the eagle owl, in arid conditions, are attracted by the abundance of prey and the presence of water (Boal et al., 2023), which may facilitate secondary AR exposure and would make these areas an indicator of the presence and distribution of ARs in the environment. It is also worth noting that, in Mediterranean semi-arid regions, watercourses are often represented by riverbeds and ravines where the presence of water is often limited or even absent. Therefore, ARs may easily concentrate in these areas due to the reduced flow and limited dispersion. In general, SGARs have lower water solubility than FGARs and degrade within a few hours when exposed to light (reviewed by Regnery et al., 2018). However, they can persist in organic matter and sediment and accumulate in the tissues of aquatic organisms (Regnery et al., 2018, 2020). Indeed, the same SGARs that were found associated with watercourses in our study were detected in faeces of Eurasian otter (Lutra lutra) and other mammals linked to riparian ecosystems from the Region of Murcia between 2020 and 2021 (Andrés-Esteso et al., 2023), confirming the relationship of these compounds with the aquatic environment. Residues of ARs in predators linked to aquatic environments have also been detected in other countries such as Germany (Regnery et al., 2024) and France

(Fournier-Chambrillon et al., 2004), suggesting that watercourses represent an important pathway for AR exposure and dissemination.

The eagle owl primarily feeds on rabbits in the study area. Therefore, given the high AR prevalence found, it is likely that this mammal frequently comes into contact with these compounds. In the Region of Murcia, as well as in other areas of Spain and the world, the rabbit is considered a pest by farmers, mainly because it directly damages various types of crops (Delibes-Mateos et al., 2018, 2020). For this reason, the rabbit is often a target species of ARs (Berny et al., 2010; Lohr and Davis, 2018), which in Spain are likely used for this purpose illegally (Colomina et al., 2024). In fact, ARs are not authorised as plant protection products, so they should not be used in agriculture and open spaces by untrained personnel. Further studies are recommended to confirm this hypothesis. Additionally, since the presence of rabbits is a key factor for the sustenance of eagle owl populations, it is critically necessary to know if the widespread presence of ARs in the environment is altering the presence of this species in the study area. It should also be noted that, in the absence of this prey species, the eagle owl can hunt other prey, including rats, which are target species for ARs and may also have contributed to the high AR prevalence found, especially in the littoral mountain ranges study site, where rabbits are scarcer.

4.2. Biomarker of effect and general health status of eagle owl nestlings

PT is considered a valid biomarker of the toxic effect that ARs cause by altering the blood's coagulative capacity (Rached et al., 2020), as this parameter changes rapidly following AR exposure in relation to the dose, and returns to baseline values after few days post-exposure (Rattner et al., 2010, 2014a). Unfortunately, PT is a species-specific parameter and reference values are not available for the eagle owl and most birds of prey (Hindmarch et al., 2019). Moreover, there is no standardisation of analysis procedures among different laboratories, partly because a specific reagent, avian thromboplastin, is required for PT analysis (Webster, 2009). This reagent is not commercially available and must be produced in the lab. All these factors make it difficult to compare results across different studies, and to perform interspecific comparisons. In our case, PT was evaluated using the same method as in samples from other nocturnal raptors in the Region of Murcia, namely the long-eared owl (Asio otus) and the barn owl (Tyto alba). In these two species, it was possible to establish a positive and significant correlation between the total concentration of ARs detected and the PT, indicating a non-acute but measurable adverse effect produced by ARs (Spadetto et al., 2024a, 2024b). In the case of eagle owl chicks, a positive but not significant relationship between the two variables was observed. This data may indicate that in this species, despite repeated exposure, nestlings had not accumulated sufficiently high hepatic AR concentrations to impair blood coagulation. Further investigations are needed to establish reference PT values in raptor species. Moreover, the standardization of laboratory protocols for analysing coagulation parameters would be critically important for assessing the immediate effects of ARs on the health of non-target wildlife.

Similarly to PT, interpreting the results obtained from the analysis of plasma biochemical parameters requires species-specific reference values. Additionally, it should be noted that these values in avian species can vary depending on the age and sex of the individuals (Agusti Montolio et al., 2018; Casado et al., 2002; Scholtz et al., 2009). The analysed biochemical parameters fall within the normal range when compared to those previously obtained in free-ranging chicks of the same species (Gómez-Ramírez et al., 2016). Some of the evaluated parameters have never been studied in the eagle owl. Where such information was unavailable, we compared the results with those obtained in nestlings or adults of other owl species (Agusti Montolio et al., 2018; Ammersbach et al., 2015; Jones and Chitty, 2020; Szabo et al., 2014).

Interestingly, the only parameters that appear higher compared to those reported for other owls are total bilirubin and magnesium (Jones and Chitty, 2020). However, bilirubin is not considered a useful marker

of liver damage in birds, due to the almost complete absence of biliverdin reductase in the liver (Jones and Chitty, 2020; Raidal, 2020), so these higher levels could be due to species and/or age differences of the subjects. Nonetheless, they are not accompanied by an alteration of other parameters indicative of liver damage. Similarly, no alterations were found in other electrolyte levels related to magnesium, indicating that the detected differences are likely interspecific. Furthermore, age is an important factor to consider since several parameters have been found to be higher in nestlings compared to juveniles/adults of the same species (Agusti Montolio et al., 2018; Gómez-Ramírez et al., 2016). Ovotransferrin was evaluated as an acute-phase protein biomarker of the activation of the immune system, as its levels increase significantly in response to infections, inflammation or trauma (Giansanti et al., 2012). It should be noted that baseline levels of ovotransferrin vary significantly depending on the species and are slightly higher in nestlings compared to adults (Horrocks et al., 2011). In the case of the eagle owl, the average found is higher than that observed in nestlings of various wild bird (non-raptor) species (Horrocks et al., 2011). However, since the rest of parameters fall within normality, we cannot state with certainty whether this is an alteration. We can therefore conclude that the eagle owl nestlings in our study were in good physiological condition. It is known that ARs can damage the liver over time and alter liver parameters (reviewed by Popov Aleksandrov et al., 2024). However, hepatic parameters were not altered, and no correlation was found between them and the total concentration of ARs detected in blood samples. Moreover, the correlation between **SARs** and glucose is weak and unlikely to be meaningful. Overall, although these individuals are exposed to ARs, the concentrations or duration of exposure were probably not sufficient to cause a detectable toxic effect.

These biochemical analysis results confirm the values reported by Gómez-Ramírez et al. (2016) and complement the panel with additional parameters that are not routinely evaluated but can be useful in research contexts or for assessing the health status of individuals admitted in wildlife rescue and rehabilitation centres.

5. Conclusions

Apex predators like the eagle owl are essential to ecosystems as they regulate prey populations, preventing resource overexploitation and maintaining balance among various species within their environment (Wallach et al., 2015). However, these species are often exposed to environmental contaminants through bioaccumulation and biomagnification along the food chain. This is particularly critical for predatory birds living in human-impacted environments, where exposure to harmful substances becomes inevitable. In our study, we found widespread AR contamination in the eagle owl, raising significant concerns for the conservation, not only of this species, but also of other predators and scavengers sharing the same habitat. Our results also suggest the potential presence and accumulation of certain ARs in the hydrological network, representing an exposure pathway that should be carefully considered, especially in semi-arid regions where water scarcity can result in reduced dilution and increased concentration of these compounds.

Although there are no clear signs of acute intoxication, chronic exposure to these compounds over time can compromise individuals' health, affecting their coagulative capacity and causing other sublethal effects. This could lead to a decrease in their biological fitness, making them more susceptible to diseases and other environmental threats (Rattner et al., 2014b). Due to the high prevalence found, further studies are needed to monitor these effects in the long term. Ultimately, this study on the eagle owl, along with others (Spadetto et al., 2024b), serves as an important starting point for further research into other top predator species such as the golden eagle (*Aquila chrysaetos*) or Bonelli's eagle (*Aquila fasciata*), which share habitat and prey and have higher protection status. In fact, to effectively conserve raptor species, it is essential to implement concrete actions aimed at mitigating the impact

of ARs and protecting the biodiversity of their ecosystems.

CRediT authorship contribution statement

Livia Spadetto: Writing - original draft, Visualization, Investigation, Formal analysis, Data curation, Conceptualization. Pilar Gómez-Ramírez: Writing – review & editing, Visualization, Supervision, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. Mario León-Ortega: Writing - review & editing, Methodology, Investigation, Conceptualization. Antonio Zamora-López: Writing - review & editing, Methodology, Investigation, Conceptualization. Sarah Díaz-García: Writing - review & editing, Methodology, Investigation, Conceptualization. José Manuel Zamora-Marín: Writing - review & editing, Methodology, Investigation, Conceptualization. Fernando Tecles-Vicente: Writing - review & editing, Methodology, Formal analysis. Luis Pardo-Marín: Writing - review & editing, Methodology, Formal analysis. José Fenoll: Writing - review & editing, Methodology, Formal analysis, Data curation. José Francisco Calvo: Methodology, Formal analysis, Data curation. Antonio Juan García-Fernández: Writing - review & editing, Visualization, Supervision, Resources, Project administration, Methodology, Investigation, Funding acquisition, Data curation, Conceptualization.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.envres.2024.120382.

Data availability

Data will be made available on request.

References

- Agusti Montolio, S., Cuenca Valera, R., Lavín González, S., Cray, C., Molina López, R., Ferron, E.S., Francisco, O.N., Marco Sánchez, I., Casas-Díaz, E., 2018. Plasma biochemistry RIs and age effect in European Strigiformes. Vet. Clin. Pathol. 47, 78–93. https://doi.org/10.1111/VCP.12559.
- Ammersbach, M., Beaufrère, H., Gionet Rollick, A., Tully, T., 2015. Laboratory blood analysis in Strigiformes-Part II: plasma biochemistry reference intervals and agreement between the Abaxis Vetscan V2 and the Roche Cobas c501. Vet. Clin. Pathol. 44, 128–140. https://doi.org/10.1111/VCP.12230.
- Andrés-Esteso, M., Jiménez, P., Spadetto, L., Castellví-Xaus, J., Méndez, A., Espín, S., García-Fernández, A.J., Gómez-Ramírez, P., 2023. Evaluación de la exposición a rodenticidas anticoagulantes en mesocarnívoros silvestres ligados a ecosistemas de

agua dulce de la Región de Murcia utilizando heces como muestras alternativas. In: XVI Congreso Internacional SECEM. Granollers (Barcelona), vol. 2023.

- Arenas-Sánchez, A., Rico, A., Vighi, M., 2016. Effects of water scarcity and chemical pollution in aquatic ecosystems: state of the art. Sci. Total Environ. 572, 390–403. https://doi.org/10.1016/J.SCITOTENV.2016.07.211.
- Badry, A., Schenke, D., Brücher, H., Chakarov, N., Grünkorn, T., Illner, H., Krüger, O., Marczak, T., Müskens, G., Nachtigall, W., Zollinger, R., Treu, G., Krone, O., 2022. Spatial variation of rodenticides and emerging contaminants in blood of raptor nestlings from Germany. Environ. Sci. Pollut. Res. 2022 (1), 1–14. https://doi.org/ 10.1007/S11356-022-20089-1.
- Badry, A., Schenke, D., Treu, G., Krone, O., 2021. Linking landscape composition and biological factors with exposure levels of rodenticides and agrochemicals in avian apex predators from Germany. Environ. Res. 193, 110602. https://doi.org/10.1016/ J.ENVRES.2020.110602.
- Bates, D., Mächler, M., Bolker, B.M., Walker, S.C., 2015. Fitting linear mixed-effects models using lme4. J. Stat. Softw. 67, 1–48. https://doi.org/10.18637/JSS.V067. 101.
- Berny, P., Esther, A., Jacob, J., Prescott, C., 2014. Risk mitigation measures for anticoagulant rodenticides as biocidal products. Final report to the European Commission (contract N°07-0307/2012/638259/ETU/D3). https://doi.org/ 10.2779/241180.
- Berny, P., Velardo, J., Pulce, C., D'Amico, A., Kammerer, M., Lasseur, R., 2010. Prevalence of anticoagulant rodenticide poisoning in humans and animals in France and substances involved. Clin. Toxicol. 48, 935–941. https://doi.org/10.3109/ 15563650.2010.533678.
- Boal, C.W., Bibles, B.D., Gicklhorn, T.S., 2023. Patterns of water use by raptors in the southern great plains. J. Raptor Res. 57, 444–455. https://doi.org/10.3356/JRR-21-70.
- Burnham, K.P., Anderson, D.R., 2002. Model Selection and Multimodel Inference: a Practical Information-Theoretic Approach, second ed. Springer, New York, NY, USA.
- Carrera, A., Navas, I., María-Mojica, P., García-Fernández, A.J., 2024. Greater predisposition to second generation anticoagulant rodenticide exposure in red foxes (*Vulpes vulpes*) weakened by suspected infectious disease. Sci. Total Environ. 907, 167780. https://doi.org/10.1016/J.SCITOTENV.2023.167780.
- Carrillo-Hidalgo, J., Martín-Cruz, B., Henríquez-Hernández, L.A., Rial-Berriel, C., Acosta-Dacal, A., Zumbado-Peña, M., Luzardo, O.P., 2024. Intraspecific and geographical variation in rodenticide exposure among common kestrels in Tenerife (Canary Islands). Sci. Total Environ. 910, 168551. https://doi.org/10.1016/J. SCITOTENV.2023.168551.
- Casado, E., Balbontin, J., Ferrer, M., 2002. Plasma chemistry in booted eagle (*Hieraaetus pennatus*) during breeding season. Comp. Biochem. Physiol. Part A Mol. Integr. Physiol. 131, 233–241. https://doi.org/10.1016/S1095-6433(01)00483-4.
- Christensen, T.K., Lassen, P., Elmeros, M., 2012. High exposure rates of anticoagulant rodenticides in predatory bird species in intensively managed landscapes in Denmark. Arch. Environ. Contam. Toxicol. 63, 437–444. https://doi.org/10.1007/ s00244-012-9771-6.
- Clauss, A., 1957. Rapid physiological coagulation method in determination of fibrinogen. Acta Haematol. 17 (4), 237–246. https://doi.org/10.1159/000205234.
- Coeurdassier, M., Fritsch, C., Jacquot, M., van den Brink, N.W., Giraudoux, P., 2018. Spatial dimensions of the risks of rodenticide use to non-target small mammals and applications in spatially explicit risk modeling. Emerg. Top. Ecotoxicol. 5, 195–227. https://doi.org/10.1007/978-3-319-64377-9.8.
- Colomina, J., Nieto, M.B., Díaz, N., María-Mojica, P., Navas, I., García-Fernández, A.J., 2024. Envenenamiento simultáneo por rodenticidas anticoagulantes de primera y segunda generación y sus implicaciones sobre la biodiversidad y salud pública: a propósito de un caso. In: III Congreso Nacional Científico de Estudiantes De Veterinaria. Murcia.
- Delibes-Mateos, M., Arroyo, B., Ruiz, J., Garrido, F.E., Redpath, S., Villafuerte, R., 2020. Conflict and cooperation in the management of European rabbit Oryctolagus cuniculus damage to agriculture in Spain. People Nat 2, 1223–1236. https://doi.org/ 10.1002/PAN3.10157/SUPPINFO.
- Delibes-Mateos, M., Farfán, M.Á., Rouco, C., Olivero, J., Márquez, A.L., Fa, J.E., Vargas, J.M., Villafuerte, R., 2018. A large-scale assessment of European rabbit damage to agriculture in Spain. Pest Manag. Sci. 74, 111–119. https://doi.org/ 10.1002/PS.4658.
- Erickson, W., Urban, D., 2004. Potential Risks of Nine Rodenticides to Birds and Nontarget Mammals: a Comparative Approach, Office of Prevention, Pesticides and Toxic Substances. United States Environmental Protection Agency, Washington, DC, USA.
- Espín, S., Andevski, J., Duke, G., Eulaers, I., Gómez-Ramírez, P., Hallgrimsson, G.T., Helander, B., Herzke, D., Jaspers, V.L.B., Krone, O., Lourenço, R., María-Mojica, P., Martínez-López, E., Mateo, R., Movalli, P., Sánchez-Virosta, P., Shore, R.F., Sonne, C., van den Brink, N.W., van Hattum, B., Vrezec, A., Wernham, C., García-Fernández, A.J., 2021. A schematic sampling protocol for contaminant monitoring in raptors. Ambio 50, 95–100. https://doi.org/10.1007/S13280-020-01341-9/ FIGURES/2.
- Espín, S., Martínez-López, E., León-Ortega, M., Calvo, J.F., García-Fernández, A.J., 2014. Factors that influence mercury concentrations in nestling Eagle Owls (*Bubo bubo*). Sci. Total Environ. 470–471, 1132–1139. https://doi.org/10.1016/J. SCITOTENV.2013.10.063.
- European Commission, 2021. Main Changes Introduced in Document N° SANTE/11312/ 2021v2 with Respect to the Previous Version (Document N° SANTE/11312/2021).
- European Environment Agency (EEA), 2018. Corine land cover (CLC) 2018 [WWW Document]. URL, Version 20. https://land.copernicus.eu/pan-european/corine -land-cover/clc2018, 3.13.24.

L. Spadetto et al.

European Union, 2016. Commission Regulation (EU) 2016/1179 of 19 July 2016 Amending, for the Purposes of its Adaptation to Technical and Scientific Progress, Regulation (EC) No 1272/2008 of the European Parliament and of the Council on Classification, Labelling and Packaging of Substances and Mixtures.

European Union, 2012. Regulation (EU) No 528/2012 of the European Parliament and of the Council of 22 May 2012 Concerning the Making Available on the Market and Use of Biocidal Products.

European Union, 2009. Directive 2009/147/EC of the European Parliament and of the Council of 30 November 2009 on the Conservation of Wild Birds.

Fourel, I., Roque, F., Orabi, P., Augiron, S., Couzi, F.-X., Puech, M.-P., Chetot, T., Lattard, V., 2024. Stereoselective bioaccumulation of chiral anticoagulant rodenticides in the liver of predatory and scavenging raptors. Sci. Total Environ. 917, 170545. https://doi.org/10.1016/J.SCITOTENV.2024.170545.

Fournier-Chambrillon, C., Berny, P.J., Coiffier, O., Barbedienne, P., Dassé, B., Delas, G., Galineau, H., Mazet, A., Pouzenc, P., Rosoux, R., Fournier, P., 2004. Evidence of secondary poisoning of free-ranging riparian mustelids by anticoagulant rodenticides in France: implications for conservation of European mink (*Mustela lutreola*). J. Wildl. Dis. 40, 688–695. https://doi.org/10.7589/0090-3558-40.4.688.

Frankova, M., Radostna, T., Aulicky, R., Stejskal, V., 2024. Less brodifacoum in baits results in greater accumulation in the liver of captive *Rattus norvegicus* in a no-choice trail. J. Pest. Sci. 1–8. https://doi.org/10.1007/S10340-023-01737-Y/FIGURES/2, 2004.

García-Fernández, A.J., María-Mojica, P., Navas, I., 2024. Avian ecotoxicology. In: Wexler, P. (Ed.), Encyclopedia of Toxicology, fourth ed., vol. 4. Elsevier, Academic Press, Oxford UK, pp. 31–43. https://doi.org/10.1016/B978-0-12-824315-2.01057-5.

Geduhn, A., Jacob, J., Schenke, D., Keller, B., Kleinschmidt, S., Esther, A., 2015. Relation between intensity of biocide practice and residues of anticoagulant rodenticides in red foxes (*Vulpes vulpes*). PLoS One 10, 1–15. https://doi.org/10.1371/journal. pone.0139191.

Giansanti, F., Leboffe, L., Pitari, G., Ippoliti, R., Antonini, G., 2012. Physiological roles of ovotransferrin. Biochim. Biophys. Acta - Gen. Subj. 1820, 218–225. https://doi.org/ 10.1016/J.BBAGEN.2011.08.004.

Gobierno de España, 2024. Confederación Hidrográfica del Segura - CHS [WWW Document]. URL https://www.chsegura.es/es/index.html, 5.5.24.

Gobierno de España, 2011. Real Decreto 139/2011, de 4 de febrero, para el desarrollo del Listado de Especies Silvestres en Régimen de Protección Especial y del Catálogo Español de Especies Amenazadas.

Gómez-Ramírez, P., Espín, S., León-Ortega, M., Botella, F., Calvo, J.F., Jiménez-Montalbán, P.J., María-Mojica, P., Martínez, J.E., Navas, I., Pérez-García, J.M., Sánchez-Zapata, J.A., Taliansky-Chamudis, A., Van den Brink, N., Martínez-López, E., García-Fernández, A., 2021. A 25 year overview of the contaminant exposure and effects in Eurasian Eagle-owl (*Bubo bubo*) from southern Spain. Airo 29, 143–165.

Gómez-Ramírez, P., Martínez-López, E., Espín, S., Jiménez, P., María-Mojica, P., Pérez-García, J.M., León-Ortega, M., García-Fernández, A.J., 2016. Haematocrit and blood biochemical parameters in free-living Eurasian eagle owls (*Bubo bubo*) from Southeastern Spain: study of age and sex differences. Eur. J. Wildl. Res. 62, 557–564. https://doi.org/10.1007/S10344-016-1028-7/TABLES/2.

Gómez-Ramírez, P., Martínez-López, E., García-Fernández, A.J., Zweers, A.J., van den Brink, N.W., 2012a. Organohalogen exposure in a Eurasian Eagle owl (*Bubo bubo*) population from Southeastern Spain: temporal–spatial trends and risk assessment. Chemosphere 88, 903–911. https://doi.org/10.1016/J. CHEMOSPHERE.2012.03.014.

Gómez-Ramírez, P., Martínez-López, E., Navas, I., María-Mojica, P., García-Fernández, A., 2012b. A modification of QuEChERS method to analyse anticoagulant rodenticides using small blood samples. Rev. Toxicol. 29, 10–14.

Gomez, E.A., Hindmarch, S., Smith, J.A., 2022. Conservation letter: raptors and anticoagulant rodenticides. J. Raptor Res. 56, 147–153. https://doi.org/10.3356/ JRR-20-122.

González-Rubio, S., Ballesteros-Gómez, A., Asimakopoulos, A.G., Jaspers, V.L.B., 2021. A review on contaminants of emerging concern in European raptors (2002–2020). Sci. Total Environ. 760, 143337. https://doi.org/10.1016/J. SCITOTENV.2020.143337.

Griminger, P., Shum, Y.S., Budowski, P., 1970. Effect of dietary vitamin K on avian brain thromboplastin activity. Poult. Sci. 49, 1681–1686. https://doi.org/10.3382/ PS.0491681.

Hindmarch, S., Rattner, B.A., Elliott, J.E., 2019. Use of blood clotting assays to assess potential anticoagulant rodenticide exposure and effects in free-ranging birds of prey. Sci. Total Environ. 657, 1205–1216. https://doi.org/10.1016/j. scitotenv.2018.11.485.

Hofstadter, D.F., Kryshak, N.F., Gabriel, M.W., Wood, C.M., Wengert, G.M., Dotters, B.P., Roberts, K.N., Fountain, E.D., Kelly, K.G., Keane, J.J., Whitmore, S.A., Berigan, W.J., Peery, M.Z., 2021. High rates of anticoagulant rodenticide exposure in California Barred Owls are associated with the wildland–urban interface. Ornithol. Appl. 123, 1–13. https://doi.org/10.1093/ORNITHAPP/DUAB036.

Horrocks, N.P.C., Irene Tieleman, B., Matson, K.D., 2011. A simple assay for measurement of ovotransferrin – a marker of inflammation and infection in birds. Methods Ecol. Evol. 2, 518–526. https://doi.org/10.1111/J.2041-210X.2011.00096. X.

Instituto Geográfico Nacional (IGN), 2016. SIOSE land cover [WWW Document]. URL. http://www.siose.es/.

Jones, M.P., Chitty, J., 2020. Raptors. Exot. Anim. Lab. Diagnosis 437–482. https://doi. org/10.1002/9781119108610.CH24.

Khidkhan, K., Yasuhira, F., Saengtienchai, A., Kasorndorkbua, C., Sitdhibutr, R., Ogasawara, K., Adachi, H., Watanabe, Y., Saito, K., Sakai, H., Horikoshi, K., Suzuki, H., Kawai, Y.K., Takeda, K., Yohannes, Y.B., Ikenaka, Y., Rattner, B.A., Ishizuka, M., Nakayama, S.M.M., 2024. Evaluation of anticoagulant rodenticide sensitivity by examining in vivo and in vitro responses in avian species, focusing on raptors. Environ. Pollut. 341, 122837. https://doi.org/10.1016/J. ENVPOL.2023.122837.

Langford, K.H., Reid, M., Thomas, K.V., 2013. The occurrence of second generation anticoagulant rodenticides in non-target raptor species in Norway. Sci. Total Environ. 450–451, 205–208. https://doi.org/10.1016/J.SCITOTENV.2013.01.100.

León-Ortega, M., 2016. Ecological Studies of Eurasian Eagle-Owl (Bubo bubo) Populations in South-Eastern Iberia: Territorial Occupancy, Reproduction, Survival, Home Range and Genetic Structure. Universidad de Murcia.

León-Ortega, M., Delgado, M. del M., Martínez, J.E., Penteriani, V., Calvo, J.F., 2016. Factors affecting survival in Mediterranean populations of the Eurasian eagle owl. Eur. J. Wildl. Res. 62, 643–651. https://doi.org/10.1007/S10344-016-1036-7/ TABLES/3.

Lohr, M.T., 2018. Anticoagulant rodenticide exposure in an Australian predatory bird increases with proximity to developed habitat. Sci. Total Environ. 643, 134–144. https://doi.org/10.1016/J.SCITOTENV.2018.06.207.

Lohr, M.T., Davis, R.A., 2018. Anticoagulant rodenticide use, non-target impacts and regulation: a case study from Australia. Sci. Total Environ. 634, 1372–1384. https:// doi.org/10.1016/J.SCITOTENV.2018.04.069.

López-Perea, J.J., Camarero, P.R., Molina-López, R.A., Parpal, L., Obón, E., Solá, J., Mateo, R., 2015. Interspecific and geographical differences in anticoagulant rodenticide residues of predatory wildlife from the Mediterranean region of Spain. Sci. Total Environ. 511C, 259–267. https://doi.org/10.1016/j. scitotenv.2014.12.042.

López-Perea, J.J., Camarero, P.R., Sánchez-Barbudo, I.S., Mateo, R., 2019. Urbanization and cattle density are determinants in the exposure to anticoagulant rodenticides of non-target wildlife. Environ. Pollut. 244, 801–808. https://doi.org/10.1016/j. envpol.2018.10.101.

Mainwaring, M.C., 2015. The use of man-made structures as nesting sites by birds: a review of the costs and benefits. J. Nat. Conserv. 25, 17–22. https://doi.org/ 10.1016/J.JNC.2015.02.007.

Martínez-Padilla, J., López-Idiáquez, D., López-Perea, J.J., Mateo, R., Paz, A., Viñuela, J., 2017. A negative association between bromadiolone exposure and nestling body condition in common kestrels: management implications for vole outbreaks. Pest Manag. Sci. 73, 364–370. https://doi.org/10.1002/PS.4435.

McClure, C.J.W., Westrip, J.R.S., Johnson, J.A., Schulwitz, S.E., Virani, M.Z., Davies, R., Symes, A., Wheatley, H., Thorstrom, R., Amar, A., Buij, R., Jones, V.R., Williams, N. P., Buechley, E.R., Butchart, S.H.M., 2018. State of the world's raptors: distributions, threats, and conservation recommendations. Biol. Conserv. 227, 390–402. https:// doi.org/10.1016/J.BIOCON.2018.08.012.

Moriceau, M.A., Lefebvre, S., Fourel, I., Benoit, E., Buronfosse-Roque, F., Orabi, P., Rattner, B.A., Lattard, V., 2022. Exposure of predatory and scavenging birds to anticoagulant rodenticides in France: exploration of data from French surveillance programs. Sci. Total Environ. 810, 151291. https://doi.org/10.1016/J. SCITOTENV.2021.151291.

Musto, C., Cerri, J., Capizzi, D., Fontana, M.C., Rubini, S., Merialdi, G., Berzi, D., Ciuti, F., Santi, A., Rossi, A., Barsi, F., Gelmini, L., Fiorentini, L., Pupillo, G., Torreggiani, C., Bianchi, A., Gazzola, A., Prati, P., Sala, G., Apollonio, M., Delogu, M., Biancardi, A., Uboldi, L., Moretti, A., Garbarino, C., 2024. First evidence of widespread positivity to anticoagulant rodenticides in grey wolves (*Canis lupus*). Sci. Total Environ. 915, 169990. https://doi.org/10.1016/J.SCITOTENV.2024.169990.

Nakayama, S.M.M., Morita, A., Ikenaka, Y., Kawai, Y.K., Watanabe, K.P., Ishii, C., Mizukawa, H., Yohannes, Y.B., Saito, K., Watanabe, Y., Ito, M., Ohsawa, N., Ishizuka, M., 2020. Avian interspecific differences in VKOR activity and inhibition: insights from amino acid sequence and mRNA expression ratio of VKORC1 and VKORC1L1. Comp. Biochem. Physiol., Part C: Toxicol. Pharmacol. 228, 108635. https://doi.org/10.1016/j.cbpc.2019.108635.

Nakayama, S.M.M., Morita, A., Ikenaka, Y., Mizukawa, H., Ishizuka, M., 2019. A review: poisoning by anticoagulant rodenticides in non-target animals globally. J. Vet. Med. Sci. 81, 298–313. https://doi.org/10.1292/jvms.17-0717.

National institute of statistics (INE) [WWW document]. URL. https://www.ine.es/. Newton, I., Shore, R.F., Wyllie, I., Birks, J.D.S., Dale, L., 1999. Empirical evidence of sideeffects of rodenticides on some predatory birds and mammals. In: Cowan, D.P., Feare, C.J. (Eds.), Advances in Vertebrate Pest Management. Furth, pp. 347–367.

O'Brien, C.S., Waddell, R.B., Rosenstock, S.S., Rabe, M.J., 2006. Wildlife use of water catchments in southwestern Arizona. Wildl. Soc. Bull. 34, 582–591, 10.2193/0091-7648(2006)34[582:WUOWCI]2.0.CO;2.

Oliva-Vidal, P., Martínez, J.M., Sánchez-Barbudo, I.S., Camarero, P.R., Colomer, M.À., Margalida, A., Mateo, R., 2022. Second-generation anticoagulant rodenticides in the blood of obligate and facultative european avian scavengers. Environ. Pollut. 315, 120385. https://doi.org/10.1016/J.ENVPOL.2022.120385.

Pérez-García, J.M., Sánchez-Zapata, J.A., Botella, F., 2012. Distribution and breeding performance of a high-density Eagle Owl *Bubo bubo* population in southeast Spain. Hous. Theor. Soc. 59, 22–28. https://doi.org/10.1080/00063657.2011.613111.

Pinheiro, J., Bates, D., Team, R.C., 2023. Nlme: linear and nonlinear mixed effects models. R package version 3, 1–162.

Popov Aleksandrov, A., Tucovic, D., Kulas, J., Popovic, D., Kataranovski, D., Kataranovski, M., Mirkov, I., 2024. Toxicology of chemical biocides: anticoagulant rodenticides – beyond hemostasis disturbance. Comp. Biochem. Physiol., Part C: Toxicol. Pharmacol. 277, 109841. https://doi.org/10.1016/J.CBPC.2024.109841.

Portal CARM Región de Murcia [WWW Document]. URL. https://www.carm.es/web/pagina?IDCONTENIDO=1&IDTIPO=180, 7.7.24.

Powolny, T., Bassin, N., Crini, N., Fourel, I., Morin, C., Pottinger, T.G., Massemin, S., Zahn, S., Coeurdassier, M., 2020. Corticosterone mediates telomere length in raptor

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chicks exposed to chemical mixture. Sci. Total Environ. 706, 135083. https://doi. org/10.1016/J.SCITOTENV.2019.135083.

- Rached, A., Moriceau, M.A., Serfaty, X., Lefebvre, S., Lattard, V., 2020. Biomarkers potency to monitor non-target fauna poisoning by anticoagulant rodenticides. Front. Vet. Sci. 7. https://doi.org/10.3389/fvets.2020.616276.
- Radeloff, V.C., Hammer, R.B., Stewart, S.I., Fried, J.S., Holcomb, S.S., McKeefry, J.F., 2005. The wildland-urban interface in the United States. Ecol. Appl. 15, 799–805. https://doi.org/10.1890/04-1413.
- Raidal, S., 2020. Laboratory diagnostics for birds. Exot. Anim. Lab. Diagnosis 429–436. https://doi.org/10.1002/9781119108610.CH23.
- Rattner, B.A., Horak, K.E., Lazarus, R.S., Goldade, D.A., Johnston, J.J., 2014a. Toxicokinetics and coagulopathy threshold of the rodenticide diphacinone in eastern screech-owls (*Megascops asio*). Environ. Toxicol. Chem. 33, 74–81. https://doi.org/ 10.1002/etc.2390.
- Rattner, B.A., Horak, K.E., Warner, S.E., Day, D.D., Meteyer, C.U., Volker, S.F., Eisemann, J.D., Johnston, J.J., 2011. Acute toxicity, histopathology, and coagulopathy in American kestrels (*Falco sparverius*) following administration of the rodenticide diphacinone. Environ. Toxicol. Chem. 30, 1213–1222. https://doi.org/ 10.1002/etc.490.
- Rattner, B.A., Horak, K.E., Warner, S.E., Johnston, J.J., 2010. Acute toxicity of diphacinone in Northern bobwhite: effects on survival and blood clotting. Ecotoxicol. Environ. Saf. 73, 1159–1164. https://doi.org/10.1016/j. ecoenv.2010.05.021.
- Rattner, B.A., Lazarus Rebecca, S., Eisenreich Karen, M., Horak Katherine, E., Volker Steven, F., Campton Christopher, M., Eisemann John, D., Meteyer Carol, U., Johnston John, J., 2012. Comparative risk assessment of the first-generation anticoagulant rodenticide diphacinone to raptors. Proc. Vertebr. Pest Conf. 25. https://doi.org/10.5070/V425110657.
- Rattner, B.A., Lazarus, R.S., Elliott, J.E., Shore, R.F., Van Den Brink, N., Brink, N. Van Den, 2014b. Adverse outcome pathway and risks of anticoagulant rodenticides to predatory wildlife. Environ. Sci. Technol. 48, 8433–8445. https://doi.org/10.1021/ es501740n.
- Rattner, B.A., Volker, S.F., Lankton, J.S., Bean, T.G., Lazarus, R.S., Horak, K.E., 2020. Brodifacoum toxicity in American kestrels (*Falco sparverius*) with evidence of increased hazard on subsequent anticoagulant rodenticide exposure. Environ. Toxicol. Chem. 39, 468–481. https://doi.org/10.1002/etc.4629.
- Regnery, J., Friesen, A., Geduhn, A., Göckener, B., Kotthoff, M., Parrhysius, P., Petersohn, E., Reifferscheid, G., Schmolz, E., Schulz, R.S., Schwarzbauer, J., Brinke, M., 2018. Rating the risks of anticoagulant rodenticides in the aquatic environment: a review. Environ. Chem. Lett. 17, 215–240. https://doi.org/10.1007/ \$10311-018-0788-6.
- Regnery, J., Rohner, S., Bachtin, J., Möhlenkamp, C., Zinke, O., Jacob, S., Wohlsein, P., Siebert, U., Reifferscheid, G., Friesen, A., 2024. First evidence of widespread anticoagulant rodenticide exposure of the Eurasian otter (*Lutra lutra*) in Germany. Sci. Total Environ. 907, 167938. https://doi.org/10.1016/j.scitotenv.2023.167938.
- Regnery, J., Schulz, R.S., Parrhysius, P., Bachtin, J., Brinke, M., Schäfer, S., Reifferscheid, G., Friesen, A., 2020. Heavy rainfall provokes anticoagulant rodenticides' release from baited sewer systems and outdoor surfaces into receiving streams. Sci. Total Environ. 740, 139905. https://doi.org/10.1016/J. SCITOTENV.2020.139905.
- Ruiz-Suárez, N., Henríquez-Hernández, L.A., Valerón, P.F., Boada, L.D., Zumbado, M., Camacho, M., Almeida-González, M., Luzardo, O.P., 2014. Assessment of anticoagulant rodenticide exposure in six raptor species from the Canary Islands (Spain). Sci. Total Environ. 485–486, 371–376. https://doi.org/10.1016/j. scitotenv.2014.03.094.
- Sánchez-Barbudo, I.S., Camarero, P.R., Mateo, R., 2012. Primary and secondary poisoning by anticoagulant rodenticides of non-target animals in Spain. Sci. Total Environ. 420, 280–288. https://doi.org/10.1016/j.scitotenv.2012.01.028.
- Sánchez-Virosta, P., León-Ortega, M., Calvo, J.F., Camarero, P.R., Mateo, R., Zumbado, M., Luzardo, O.P., Eeva, T., García-Fernández, A.J., Espín, S., 2020. Blood

concentrations of 50 elements in Eagle owl (*Bubo bubo*) at different contamination scenarios and related effects on plasma vitamin levels. Environ. Pollut. 265, 115012. https://doi.org/10.1016/J.ENVPOL.2020.115012.

- Scholtz, N., Halle, I., Flachowsky, G., Sauerwein, H., 2009. Serum chemistry reference values in adult Japanese quail (*Coturnix coturnix japonica*) including sex-related differences. Poult. Sci. 88, 1186–1190. https://doi.org/10.3382/PS.2008-00546.
- Shenbrot, G., Krasnov, B., Burdelov, S., 2010. Long-term study of population dynamics and habitat selection of rodents in the Negev Desert. J. Mammal. 91, 776–786.
- https://doi.org/10.1644/09-MAMM-S-162.1/2/JMAMMAL-91-4-776-FIG2.JPEG.
 Shore, R.F., Taggart, M.A., 2019. Population-level impacts of chemical contaminants on apex avian species. Curr. Opin. Environ. Sci. Heal. 11, 65–70. https://doi.org/ 10.1016/J.COESH.2019.06.007.
- Silveira, G., Frair, J.L., Murphy, L., Ellis, J.C., Needle, D., Cunningham, S.A., Watson, A., Facka, A., Tate, P., Webb, S., Royar, K., Bernier, C., Keller, T., Schuler, K., 2024. Drivers of anticoagulant rodenticide exposure in Fishers (*Pekania pennanti*) across the northeastern United States. Front. Ecol. Evol. 12, 1304659. https://doi.org/ 10.3389/FEVO.2024.1304659.
- Spadetto, L., García-Fernández, A.J., Zamora-López, A., Zamora-Marín, J.M., León-Ortega, M., Tórtola-García, M., Tecles-Vicente, F., Fenoll-Serrano, J., Cava-Artero, J., Calvo, J.F., Gómez-Ramírez, P., 2024a. Comparing anticoagulant rodenticide exposure in barn owl (*Tyto alba*) and common kestrel (*Falco tinnunculus*): a biomonitoring study in an agricultural region of southeastern Spain. Environ. Pollut. 362, 124944. https://doi.org/10.1016/J.ENVPOL.2024.124944.
- Spadetto, L., Gómez-Ramírez, P., Zamora-Marín, J.M., León-Ortega, M., Díaz-García, S., Tecles, F., Fenoll, J., Cava, J., Calvo, J.F., García-Fernández, A.J., 2024b. Active monitoring of long-eared owl (*Asio otus*) nestlings reveals widespread exposure to anticoagulant rodenticides across different agricultural landscapes. Sci. Total Environ. 918, 170492. https://doi.org/10.1016/J.SCITOTENV.2024.170492.

Spanish national agency of Meteorology - AEMET [WWW document]. URL. htt ps://www.aemet.es/, 6.15.24.

- Steward, A.L., Von Schiller, D., Tockner, K., Marshall, J.C., Bunn, S.E., 2012. When the river runs dry: human and ecological values of dry riverbeds. Front. Ecol. Environ. 10, 202–209. https://doi.org/10.1890/110136.
- Szabo, Z., Klein, A., Jakab, C., 2014. Hematologic and plasma biochemistry reference intervals of healthy adult barn owls (*Tyto alba*). Avian Dis. 58 (1), 228–231. https:// doi.org/10.1637/10715-111013-REG.
- Thomas, P.J., Mineau, P., Shore, R.F., Champoux, L., Martin, P.A., Wilson, L.K., Fitzgerald, G., Elliott, J.E., 2011. Second generation anticoagulant rodenticides in predatory birds: probabilistic characterisation of toxic liver concentrations and implications for predatory bird populations in Canada. Environ. Int. 37, 914–920. https://doi.org/10.1016/j.envint.2011.03.010.
- Vicedo, T., Navas, I., María-Mojica, P., García-Fernández, A.J., 2024. Widespread use of anticoagulant rodenticides in agricultural and urban environments. A menace to the viability of the endangered Bonelli's eagle (*Aquila fasciata*) populations. Environ. Pollut. 358, 124530. https://doi.org/10.1016/J.ENVPOL.2024.124530.
- Wallach, A.D., Ripple, W.J., Carroll, S.P., 2015. Novel trophic cascades: apex predators enable coexistence. Trends Ecol. Evol. 30, 146–153. https://doi.org/10.1016/j. tree.2015.01.003.
- Webster, K.H., 2009. Validation of a Prothrombin Time (PT) Assay for Assessment of Brodificoum Exposure in Japanese Quail and Barn Owls. Simon Fraser University, Canada.
- Williams, D.D., 2005. The Biology of Temporary Waters, vol. 2005. Oxford Academic, Oxford. https://doi.org/10.1093/acprof:oso/9780198528128.001.0001, 1 Sept. 2007.
- Zamora-Marín, J.M., Zamora-López, A., Jiménez-Franco, M.V., Calvo, J.F., Oliva-Paterna, F.J., 2021. Small ponds support high terrestrial bird species richness in a Mediterranean semiarid region. Hydrobiologia 848, 1623–1638. https://doi.org/ 10.1007/S10750-021-04552-7/METRICS.